



Evaluation of Empirical Functions and Fate of Isomaltose

**Toluwase Hezekiah Fatoki^{1*}, David Morakinyo Sanni¹,
Daniel Uwaremhevho Momodu², Harriet Unuagbon Ugboko³,
Catherine Joke Adeseke¹ and Bolanle Christianah Faleye⁴**

¹*Department of Biochemistry, Federal University of Technology, PMB 704, Akure, Ondo State, Nigeria.*

²*Department of Industrial Chemistry, Federal University Oye, PMB 373, Oye, Ekiti State, Nigeria.*

³*Department of Biological Sciences, Covenant University, PMB 1023, Ota, Ogun State, Nigeria.*

⁴*Department of Chemical Science, Joseph Ayo Babalola University, Ikeji-Arakeji, PMB 5006, Ilesa, Osun State, Nigeria.*

Authors' contributions

This work was carried out in collaboration between all authors. Author THF designed the study, performed the computational analysis, wrote the protocol and wrote the first draft of the manuscript. Authors DMS and DUM managed the analyses of the study. Authors HUU, CJA and BCF managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JALSI/2018/39370

Editor(s):

(1) Palanisamy Arulselvan, Institute of Bioscience, Universiti Putra Malaysia, Malaysia.

Reviewers:

(1) César Luiz Da Silva Guimarães, Interamerican University of Rondônia, Brazilian Institute of Environment and of Natural Renewable Resources, Brazil.

(2) Keagile Bati, University of Botswana, Botswana.

(3) Nelson Pérez Guerra, University of Vigo, Spain.

(4) Oguz Ozcelik, Firat University, Turkey.

Complete Peer review History: <http://www.sciencedomain.org/review-history/23105>

Original Research Article

Received 13th November 2017

Accepted 4th February 2018

Published 9th February 2018

ABSTRACT

Aim: To validate the empirical functions of isomaltose and proposed its fates in metabolism using computational analysis.

Place and Duration of Study: This work was carried out at the Department of Biochemistry at the Federal University of Technology Akure Nigeria in 2017.

Methodology: We make use of text mining of experimental articles on isomaltose, predicted the potential targets using *Swisstargetprediction* server, and the *SwissSimilarity* server was used for drug similarity analysis.

*Corresponding author: E-mail: hezekiahfatoki@gmail.com;

Results: Isomaltose exhibits antimicrobial activity, biodegradability, non-toxic, and find its use in food, biochemistry and pharmaceutical industries as a prebiotic molecule, precursor metabolite and potent medicine respectively. Isomaltose possibly has a lower glycemic index, with the fate that implicated the production of diglucosamine, by the help of specific aminotransferase, and that there is a possibility of biosynthesis of secondary metabolites through isomaltose pathway.

Conclusion: This study provides the future areas of research and application of isomaltose.

Keywords: Isomaltose; diglucosamine; novel pathway; *in silico*; functions; fate.

1. INTRODUCTION

Isomaltose (6- O- α -D-glucopyranosyl-D-glucopyranose) is a disaccharide that comprises an alpha-1,6-glycosidic bond between two glucose molecules. Isomaltose is a member of the isomalto-oligosaccharide group, which are known to improve the general well-being of humans and animals when ingested orally daily at a moderate dose of 8-10 g, with an average of 2.8 mg availability in the urine of healthy, fasting individual [1,2,3]. Isomaltose caloric value has been found to be less than 50% that of sucrose, and its viscosity is lower than that of maltose [4]. Although isomaltose is actively hydrolysed by the disaccharidase (isomaltase) in the small intestinal mucosa, small amounts of the unhydrolyzed may be absorbed and excreted in the urine when high intestinal concentrations are attained. However, congenital sucrase-isomaltase deficiency (CSID), an autosomal recessive disaccharidase deficiency, has been recognised as the cause of isomaltose intolerance [5]. From a practical point of view, isomaltose is used as a prototype of the starch and glycogen branching, once branching in these polymers occurs through the (α -1,6) glycosidic bond between two glucose residues of the polymeric chain [6]. Surprisingly, despite the importance of the isomaltose, few published studies can be found that provide insights into its metabolic functions. In this study, functions of isomaltose are propounded and its fates in metabolism are predicted by *in silico* analysis. This study provides an advanced knowledge of isomaltose for possible industrial applications.

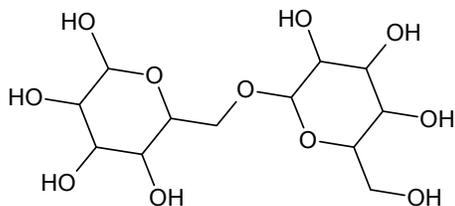


Fig. 1. Typical structure of isomaltose

2. MATERIALS AND METHODS

2.1 Text Mining of Functional Properties

The metabolic pathway in an organism where isomaltose is located was obtained from www.kegg.jp. The text mining for 'isomaltose' was done by queried PubMed database and other texts were obtained from Google search. Reports on various functions of isomaltose were intelligently mined from the available literatures, and possible scientific bases were postulated for the functions and fate of isomaltose.

2.2 *In silico* Target Prediction

The potential target for isomaltose and its proposed associated metabolites, were analysis using *Swisstargetprediction* server (<http://www.swisstargetprediction.ch>) using the SMILES (Simplified Molecular Input Line Entry Specification) obtained through Chemsketch (<http://www.acdlabs.com>), and *Homo sapiens* was selected as the target source [7].

2.3 Drug Similarity Search

Isomaltose and its proposed associated metabolites were compared to the available US Food and Drug Agency (FDA) approved drug database by combine screening method using *SwissSimilarity* server (<http://www.swissSimilarity.ch>) [8].

3. RESULTS AND DISCUSSION

At the time of this study, PubMed journal database has 564 published articles that included 'isomaltose' between the year 1949 and 2017, while only 20 of these articles make use of 'isomaltose' in their title. Also, there are 47 published articles that include 'diglucosamine', and only 1 of the articles make use of 'diglucosamine' in its title.

3.1 Isomaltose as a Prebiotic Molecule

Isomalto-oligosaccharide (IMO) is a non-digestible, low calorie health sweetener that supports the proliferation of the beneficial bacteria residing in the large intestine (colon), and consequently acts as a prebiotic [9,10]. Prebiotic is defined as a non-digestible food ingredient that beneficially affects the host by selectively stimulating the growth and/or activity of one or a limited number of bacteria in the colon, that can improve the host health [11-14]. However, the degree of non-digestibility must be established. In the case of isomaltose, active hydrolysis is balanced with non-hydrolysis by glucose homeostasis which reflects the balance between the rate of glucose entering the circulation and the rate of glucose leaving the circulation.

Based on the differences observed in the glycemic index of sucrose and isomaltulose, that ingestion of slowly absorbed isomaltulose attenuated postprandial hyperglycemia by reducing oral glucose appearance by secretion of glucagon-like peptide 1 (GLP-1), inhibiting endogenous glucose production (EGP), and increasing splanchnic glucose uptake (SGU) compared with ingestion of rapidly absorbed sucrose in patients with type 2 diabetes mellitus (T2DM) [15]. It could be deduced that isomaltose will also have lower glycemic index than maltose or sucrose, in that both isomaltulose (α -D-glucopyranosyl-1,6-D-fructofuranose) and isomaltose, are catalyzed by isomaltase. Thus, isomaltose will be a low glycemic index carbohydrate.

3.2 Isomaltose as a Precursor Metabolite

Previous study has shown that natural and engineered bacteria can effectively assimilate isomaltose by implication of phospho- α -glucosidase and phosphoenolpyruvate-dependent maltose phosphotransferase system (PEP:PTS), leading to the formation of isomaltose-6-phosphate [16,17]. In *Streptomyces coelicolor*, fructose, N-acetylglucosamine, and possibly two other carbohydrates are transported by the phosphotransferase system [18,19]. This reveals possible association of isomaltose with glucosamine pathway. Different aminoglycoside antibiotics has been found and produced by few species of *Streptomyces*, which include paromomycin, trobamycin, kanamycin and framycetin (Table 2).

Glucosamine (GlcN) is one of the most commonly fermented substrates among human and animal *Bifidobacteria* [20], and numerous *Bacteroides spp.* found in the colon are known to ferment GlcN [21].

However, from a biological point of view, the presence of isomaltose has been associated with metabolic and renal illness [3,22], which include age-related macular degeneration (AMD), a leading disease that cause vision loss and blindness at old age [23]. This may be the result of inhibition of arginine:isomaltose aminotransferase (AIAT), a possible novel enzyme that catalyzes formation of diglucosamine from isomaltose. This inhibition could be the cause of high blood plasma concentrations of isomaltose and associated amino acids such as phenylalanine, tyrosine, methionine, while lower level of N1-Methyl-2-pyridone-5-carboxamide, biliverdin and hydrocortisone could be an indication of blockage of purine alkaloid route, and diversion to alternative route for biosynthesis of alkaloids derived from ornithine, lysine and nicotinic acid; and reduction in iron absorption, as implicated in affected metabolic pathways [23]. Previous report has shown that glucosamine sulfate inhibits tumor necrosis factor alpha (TNF- α), interferon gamma (IFN- γ), and proinflammatory cytokine-induced production of Intercellular Adhesion Molecule 1 (ICAM-1) in human retinal pigment epithelial cells *in vitro* [24,25]. Glucosamine sulfate (GlcN-6S) is essentially beneficial in the treatment of osteoarthritis [26], and it is possibly produced from GlcN in the presence of dietary methionine [27] or from the degradation of heparitin sulfate and chondroitin sulfate by the specific glucosamine-6-O-sulphatase in some bacteria [28].

In addition, IMO increases the production of beneficial short-chain fatty acids such as butyrate and the absorption of calcium and magnesium [29,30], in that they serve as backbone for the formation of lipid A. Diglucosamine (GlcN₂) is an isomaltose-like compound formed from two units of glucosamine (2-amino-2-deoxyglucose) or by hydrolysis of chitin. The occurrence of diglucosamine (Fig. 3) has been reported present as the component of the lipid A in the membrane of some bacteria, where it played vital role in protection against environmental stresses [31,32]. No evidence of GlcN-related effects on the use of oral GlcN for individuals at risk for diabetes, or those with type 1 or 2 diabetes, or normo-glycemic with respect to any adverse

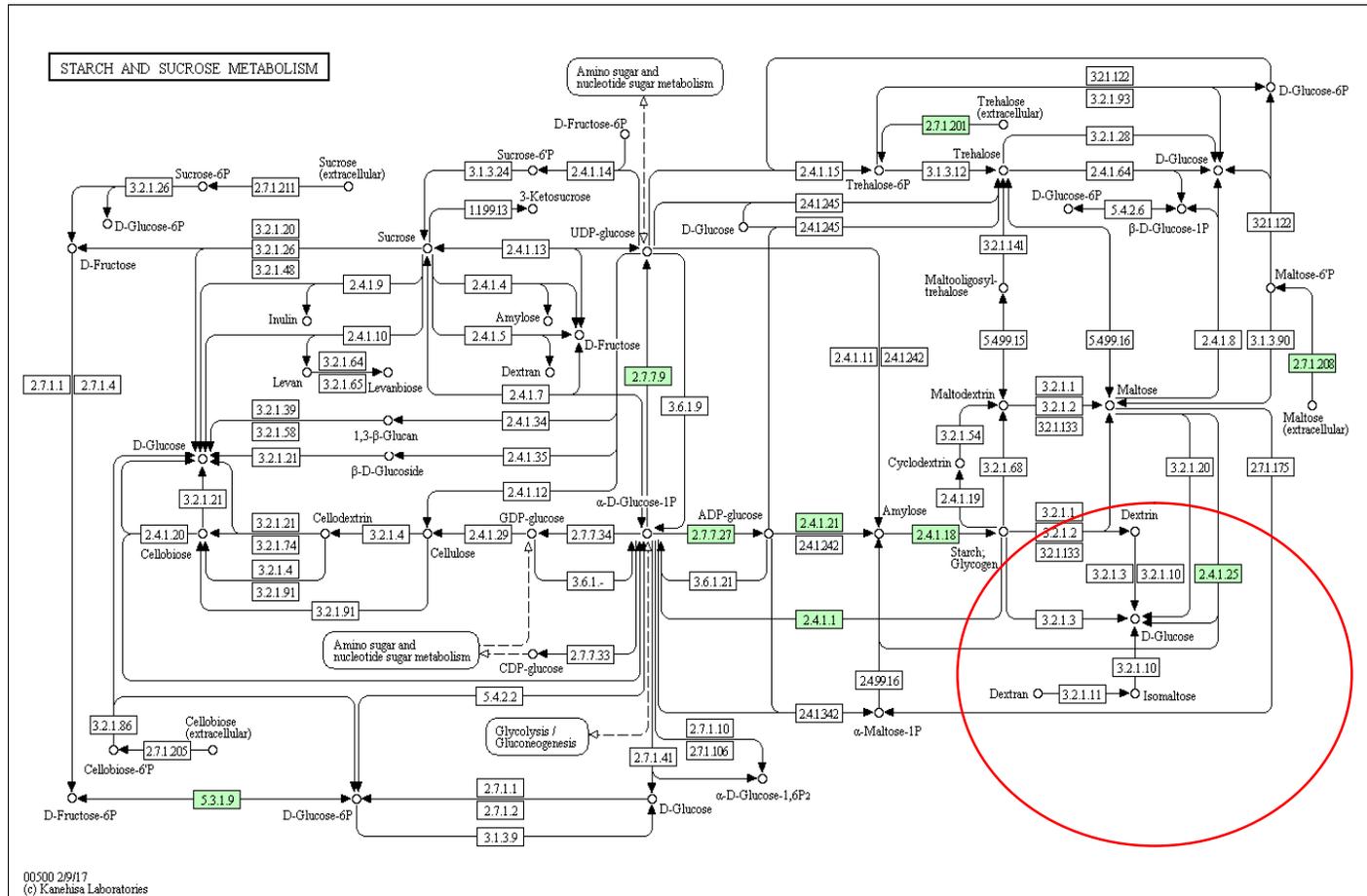


Fig. 2. Starch and sucrose metabolic pathways obtained from www.kegg.jp. Area in red shows the position of isomaltose

effects on sugar metabolism [33]. GlcN acts as a preferred substrate for the biosynthesis of glycosaminoglycan chains and subsequently to produce aggrecan and other proteoglycans of cartilage [26]. In humans, the average endogenous production of glucosamine is 12 g/day [34]. This may lead to the possible fate of isomaltose in metabolism, as it could serve as salvage pathway to the production of GlcN₂ and eventually GlcN, with the help of specific aminotransferase and glycoside hydrolase sequentially as shown in Fig. 4. A typical aminotransferase is found the cytosol while the glycoside hydrolase is in the lysosome.

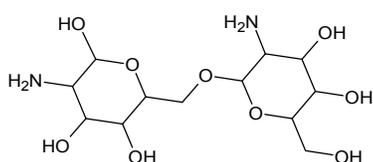


Fig. 3. Typical structure of diglucosamine

3.3 Isomaltose as a Potent Medicine

Alkyl glycosides are a group of attractive surfactants. They exhibit antimicrobial activity,

biodegradability and low toxicity [35], and find their use in pharmaceutical industry [36]. Isomaltose is one of the novel alkylated halogenated disaccharides which were patented as a formulation, based on the pharmaceutical efficacy such as permeation enhancer and antimicrobial activity [37]. Study has shown that the entrapment of bovine serum albumin (BSA) into the liposomes was increased by the GlcN₂ [38]. Isomalto-oligosaccharides have been used for treatment of chronic constipation and hyperlipidemia occurring as complications of maintenance haemodialysis [2]. In *Aspergillus nidulans* and *Aspergillus oryzae*, study has shown that isomaltose is a physiological inducer for amylase [39,40], and that inhibition of adenylosuccinate synthase (ADSS) or a more proximal enzyme in the adenylosuccinate (S-AMP) biosynthesis pathway, adenylosuccinate lyase (ADSL), lowers S-AMP levels and impairs glucose-stimulated insulin secretion (GSIS) [41]. Also, melibiose and isomaltose, were reported as a promoter of quercetin glycoside absorption in rats by increasing glycoside hydrolysis in the intestinal lumen, and that α-1,6 linkage is involved in this process [42].

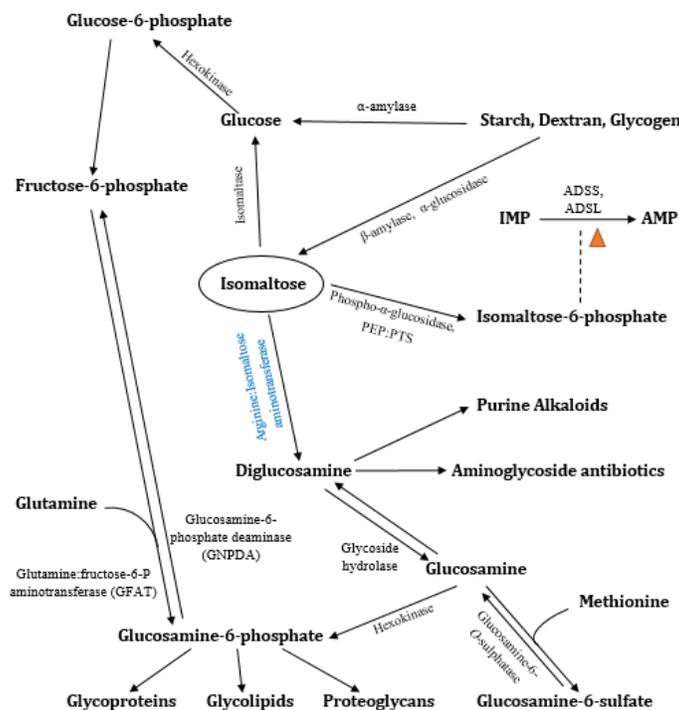


Fig. 4. Possible fate of isomaltose in metabolism. The blue colour indicates proposed enzyme (arginine:isomaltose aminotransferase) while orange colour indicates induction of adenylosuccinate synthase (ADSS) and adenylosuccinate lyase (ADSL) by isomaltose-6-phosphate

Table 1. Potential targets of isomaltose and its proposed associated metabolites

Target	Uniprot ID	Associated compounds			
		A	B	C	D
Cyclin-dependent kinase 1,2,3,4,6	P06493, P24941, Q00526, P11802, Q00534	****		**	
Fibroblast growth factor 1, 2	P05230, P09038	****		**	
Muscarinic acetylcholine receptor M1,2,3,4,5	P11229, P08172, P20309, P08173, P08912	***			
Vascular endothelial growth factor A	P15692	***			
Heparanase 8 kDa subunit	Q9Y251	**			
Inactive heparanase-2	Q8WWQ2	**			
Cation-independent mannose-6-phosphate receptor	P11717		**		*
Pyruvate kinase isozymes M1/M2	P14618		*		*
Microtubule-associated protein tau	P10636			***	
Tissue and plasma alpha-L-fucosidase	P04066, Q9BTY2			**	
Glucoamylase	O43451			*	
Sucrase	P14410			*	
Carbonic anhydrase 1, 12	P00915, O43570			*	

* (20-30%), ** (40-50%), *** (50-60%), **** (70-80%) Probability on Target. Probabilities have been computed based on a cross-validation. They may therefore not represent the actual probability of success for any new molecule.

A = Isomaltose, C(C1C(C(C(C(O1)OCC2C(C(C(C(O2)O)O)O)O)O)O)O)O)O

B = Isomaltose-6-phosphate, OC1OC(COC2OC(COP(O)(O)=O)C(O)C(O)C2O)C(O)C(O)C1O

C = Diglucosamine, NC1C(O)OC(COC2OC(CO)C(O)C(O)C2N)C(O)C1O

D = Glucosamine-6-phosphate, C(C1C(C(C(C(O1)O)N)O)O)OP(=O)(O)O

Table 2. Similarity data for isomaltose and its proposed associated metabolites

FDA approved drug	Drugbank ID	Similarity score	
		Isomaltose	Diglucosamine
Kanamycin	DB01172	0.300	0.860
Paromomycin	DB01421	0.346	0.810
Lactulose	DB00581	0.994	0.754
Tobramycin	DB00684	0.159	0.679
Glucosamine	DB01296	0.205	0.605
Neomycin	DB00994	0.207	0.605
Framycetin	DB00452	0.207	0.605
Gentamycin	DB00798	-	0.267

Therefore, there is possibility of biosynthesis of secondary metabolites through isomaltose pathway (Fig. 4), especially aminoglycoside antibiotics and rare alkaloids that consist of adenine-like molecule, which could help in cellular functioning such as pathogen exocytosis, and proper protein folding by preventing aggregation [43]. It could be said that if isomaltose is an intermedia, then GlcN₂ will likely be an obligatory intermedia in the synthesis of specific purine alkaloid. Intermedia is a compound which can be the final product of any pathway. An obligatory intermedia is a compound which follows the intermedia in the synthesis process of the alkaloid and metabolism pathway. Moreover, inosine monophosphate (IMP) is an intermedia whenever purine is an alkaloid precursor. For non-amino acid precursor, the obligatory intermedia is derived either from intermedia by S-Adenosylmethionine-dependent N-methylation or by a coupling reaction [44].

Based on the data of Tables 1 and 2, isomaltose possibly has physiological induction property, due to its action on cyclin-dependent kinases, and fibroblast growth factor among others. These two targets are associated with heat shock protein Hsp70 and Hsp90 respectively. The highest similarity score is 1. The high similarity match of isomaltose to lactulose (a synthetic disaccharide used in the treatment of constipation and hepatic encephalopathy, as well as in the diagnosis of gastrointestinal disorders), confirmed the earlier empirical function of isomaltose [2]. Diglucosamine showed more pharmacological properties for neurodegenerative dementia such as Parkinson disease, due to its probable action on microtubule-associated protein tau. It has high similarity score to antibiotics drugs (kanamycin, paromomycin, tobramycin, neomycin, and framycetin), in addition to lactulose. This suggests the possibility of diglucosamine being a metabolically active product of isomaltose, and

provide platform for synthetic development of novel drug molecules [45].

4. CONCLUSION

This study revealed the potentials of isomaltose as a material for pharmaceutical, food and biochemistry applications. The major problem initiated by this study is the occurrence of arginine: isomaltose aminotransferase (AIAT) and the pathway leading to alkaloid biosynthesis. However, there is an urgent need for further research to validate and provide biochemical solution to these problems.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Lee HC, Kim JH, Kim SY, Lee JK. Isomaltose production by modification of the fructose-binding site on the basis of the predicted structure of sucrose isomerase from "*Protaminobacter rubrum*". Applied and Environmental Microbiology. 2008; 5183–5194. DOI: 10.1128/AEM.00181-08
2. Goulas AK, Fisher DA, Grimble GK, Grandison AS, Rastall RA. Synthesis of isomaltooligosaccharides and oligodextrans by the combined use of dextransucrase and dextranase. Enzyme and Microbial Technology. 2004;35:327–338.
3. Vitek V, Vitek K, Lin H, Cowley RA. Isomaltose excretion in health, severe injury, and disease. Metabolism. 1972;21(8):701-712.
4. Harrion MD, Purdue JC, Hoffman AJ. Food products comprising a slowly digestible or

- digestion resistant carbohydrate composition. U.S. patent 2007/0184177 A1; 2007.
5. Cohen S, Stanley. The clinical consequences of sucrase-isomaltase deficiency. *Molecular and Cellular Pediatrics*. 2016;3(5):1-4.
DOI: 10.1186/s40348-015-0028-0
 6. Javaroni F, Ferreira ABB, da Silva CO. The (α -1,6) glycosidic bond of isomaltose: A tricky system for theoretical conformational studies. *Carbohydrate Research*. 2009;344:1235–1247.
DOI: 10.1016/j.carres.2009.04.022
 7. Gfeller D, Michelin O, Zoete V. Shaping the interaction landscape of bioactive molecules. *Bioinformatics*. 2013;29:3073–3079.
 8. Zoete V, Daina A, Bovigny C, Michelin O. SwissSimilarity: A web tool for low to ultra-high throughput ligand-based virtual screening. *Journal of Chemical Information and Modeling*. 2016;56(8):1399–1404.
 9. Rycroft CE, Jones MR, Gibson GR, Rastall RA. A comparative in vitro evaluation of the fermentation properties of prebiotic oligosaccharide. *Journal of Applied Microbiology*. 2001;91:878-87.
 10. Kohmoto T, Fukui F, Takaku H, Machida Y, Arai M, Mitsuoka T. Effect of isomaltoligosaccharides on human fecal flora. *Bifidobacteria Microflora*. 1988;7:61-69.
 11. Gibson GR, Roberfroid MB. Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. *Journal of Nutrition*. 1995; 125(6):1401-12.
 12. Gibson GR, Probert HM, Van Loo J, Rastall RA, Roberfroid MB. Dietary modulation of the human colonic microbiota: Updating the concept of prebiotics. *Nutrition Research Reviews*. 2004;17:259–275.
 13. Roberfroid MB. Prebiotics: The concept revisited. *Journal of Nutrition*. 2007;137: 830S–837S.
 14. Chow J. Probiotics and prebiotics: A brief overview. *Journal of Renal Nutrition*. 2002;12(2):76-86.
 15. Ang Meidjie, Linn Thomas. Comparison of the effects of slowly and rapidly absorbed carbohydrates on postprandial glucose metabolism in type 2 diabetes mellitus patients: A randomized trial. *American Journal of Clinical Nutrition*. 2014;100: 1059–68.
DOI: 10.3945/ajcn.113.076638
 16. Thompson J, Robrish SA, Immel S, Lichtenthaler FW, Hall BG, Pikis A. Metabolism of sucrose and its five linkage-isomeric -D-Glucosyl-D-fructoses by *Klebsiella pneumoniae*. *Journal of Biological Chemistry*. 2001;276(40): 37415–37425.
 17. Abe K, Kuroda A, Takeshita R. Engineering of *Escherichia coli* to facilitate efficient utilization of isomaltose and panose in industrial glucose feedstock. *Applied Microbiology and Biotechnology*. 2017;101:2057–2066.
DOI: 10.1007/s00253-016-8037-z
 18. Nothhaft H, Dresel D, Willimek A, Mahr K, Niederweis M, Titgemeyer F. The phosphotransferase system of *Streptomyces coelicolor* is biased for *N*-acetylglucosamine metabolism. *Journal of Bacteriology*. 2003;185:7019–7023.
 19. Bertram R, Schlicht M, Mahr K, Nothhaft H, Saier MH, Titgemeyer F. *In silico* and transcriptional analysis of carbohydrate uptake systems of *Streptomyces coelicolor* A3(2). *Journal of Bacteriology*. 2004;186(5):1362–1373.
DOI: 10.1128/JB.186.5.1362–1373.2004
 20. Crociani F, Alessandrini A, Mucci MM, Biavati B. Degradation of complex carbohydrates by *Bifidobacterium* spp. *International Journal of Food Microbiology*. 1994;24:199–210.
 21. Salyers AA, Vercellotti JR, West SEH, Wilkins TD. Fermentation of mucin and plant polysaccharides by strains of bacteroides from the human colon. *Applied Environmental Microbiology*. 1977;33:319–322.
 22. Butscheid M, Schafer C, Brenner S, Alscher D, Murdter T, Niwa T, Frischmann M, Pischetsrieder M, Klotz U. Unchanged serum levels of advanced glycation endproducts in patients with liver disease. *Naunyn Schmiedebergs Archive of Pharmacology*. 2007;375(6):401-6.
 23. Luo D, Deng T, Yuan W, Deng H, Jin M. Plasma metabolomic study in Chinese patients with wet age-related macular degeneration. *BMC Ophthalmology*. 2017;17:165.
DOI: 10.1186/s12886-017-0555-7
 24. Chen JT, Liang JB, Chou CL, Chien MW, Shyu RC, Chou PI, Lu DW. Glucosamine sulfate inhibits TNF-alpha and IFN-gamma-induced production of ICAM-1 in

- human retinal pigment epithelial cells *in vitro*. Investigative Ophthalmology & Visual Science. 2006a;47(2):664-72.
25. Chen JT, Chen CH, Horng CT, Chien MW, Lu DW, Liang JB, Tai MC, Chang YH, Chen PL, Chen YH. Glucosamine sulfate inhibits proinflammatory cytokine-induced icam-1 production in human conjunctival cells *in vitro*. Journal of Ocular Pharmacology and Therapeutics. 2006b; 22(6):402-16.
 26. Reginster JY, Bruyere O, Neuprez A. Current role of glucosamine in the treatment of osteoarthritis. Rheumatology. 2007;46:731–735.
DOI: 10.1093/rheumatology/kem026
 27. Yamagishi Y, Igarashi M, Suzuki A, Suguro S, Hirano S, Nagaoka I. Evaluation of the effect of methionine and glucosamine on adjuvant arthritis in rats. Experimental and Therapeutic Medicine. 2012;4:640-644.
DOI: 10.3892/etm.2012.668
 28. Bruce JS, McLean MW, Williamson FB, Long WF. *Flavobacterium heparinum* 6-*O*-sulphatase for N-substituted glucosamine 6-*O*-sulphate. FEBS European Journal of Biochemistry. 1985;152:75-82.
 29. Kashimura J, Kimura M, Itokawa Y. The effects of isomaltulose, isomalt and isomaltulose-based oligomers on mineral absorption and retention. Biological Trace Element Research. 1996;54(3):239-50.
 30. Kashimura J, Kimura M, Itokawa Y. The effects of isomaltulose-based oligomers feeding and calcium deficiency on mineral retention in rats. Journal of Nutritional Science and Vitaminology. 1996;42(1):69-76.
 31. Robert CB, Thomson M, Vercellone A, Gardner F, Ernst RK, Larrouy-Maumus G, Nigou J. Mass spectrometry analysis of intact *Francisella* bacteria identifies lipid A structure remodeling in response to acidic pH stress. Biochimie. Epub. 2017;141:16-20.
DOI: 10.1016/j.biochi.2017.08.008
 32. Lamarche MG, Wanner BL, Cre ´pin S, Harel J. The Pho regulon and bacterial virulence: a regulatory network connecting phosphate homeostasis and pathogenesis. FEMS. Microbiology Review. 2008;32: 461–473.
 33. Simon RR, Marks V, Leeds AR, Anderson JW. A comprehensive review of oral glucosamine use and effects on glucose metabolism in normal and diabetic individuals. Diabetes Metabolism Research and Reviews. 2011;27:14–27.
DOI: 10.1002/dmrr.1150
 34. Anderson JW, Nicolosi RJ, Borzelleca JF. Glucosamine effects in humans: A review of effects on glucose metabolism, side effects, safety considerations and efficacy. Food and Chemical Toxicology. 2005;43:187–201.
 35. Matsumura S, Imai K, Yoshikawa S, Kawada K, Uchibor T. Surface activities, biodegradability and antimicrobial properties of n-alkyl glucosides, mannosides and galactosides. Journal of the American Oil Chemists’ Society. 1990;67:996–1001.
 36. von Rybinski W, Angew KH. Alkyl polyglycosides – properties and applications of a new class of surfactants. Angewandte Chemie International Edition. 1998;37:1328–1345.
 37. Memarzadeh Bahram. Halogenated alkyl di- and trisaccharides, pharmaceutical formulations, diagnostic kits and methods of treatment. US Patent 2008/0171721 A1; 2008.
 38. Murakami S, Ono T, Sakai S, Ijima H, Kawakami K. Effect of Diglucosamine on the entrapment of protein into liposomes. Journal of Liposome Research. 2006;16:103–112.
DOI: 10.1080/08982100600680667
 39. Kato N, Murakoshi Y, Kato M, Kobayashi T, Tsukagoshi N. Isomaltose formed by α -glucosidases triggers amylase induction in *Aspergillus nidulans*. Current Genetics. 2002;42:43-50.
DOI: 10.1007/s00294-002-0325-8
 40. Tonomura K, Suzuki H, Nakamura N, Kuraya K, Tanabe O. On the inducers of α -amylase formation in *Aspergillus oryzae*. Agricultural and Biological Chemistry. 1961;25:1-6.
 41. Gooding JR, Jensen MV, Dai X, Wenner BR, Lu D, Arumugam R, Ferdaussi M, MacDonald PE, Newgard C. Adenylosuccinate is an insulin secretagogue derived from glucose-induced purine metabolism. Cell Reports. 2015;13:157–167.
DOI: 10.1016/j.celrep.2015.08.072
 42. Tanaka S, Shinoki A, Hara H. Melibiose, a non-digestible disaccharide, promotes

- absorption of quercetin glycosides in rat small intestine. *Journal of Agricultural and Food Chemistry*. 2016;1-36.
DOI: 10.1021/acs.jafc.6b03714
43. Sanni DM, Fatoki TH, Kolawole AO, Akinmoladun AC. Xeronine structure and function: Comparative mastery of its mystery. *In silico Pharmacology*. 2017;5: 8.
DOI: 10.1007/s40203-017-0028-y
44. Aniszewski Tadeusz. Alkaloids – secrets of life: Alkaloid chemistry, biological significance, applications and ecological role. Elsevier, Netherlands. 2007;61-139.
45. Xu G, Wang X, Zhang M, Zeng L, Li H, Gao P, Zhang J. Design, synthesis, and evaluation of glycopeptides containing glucosamine. *Medicinal Chemistry*. 2014;4(5):494-500.
DOI: 10.4172/2161-0444.1000184

© 2018 Fatoki et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
<http://www.sciencedomain.org/review-history/23105>